

Miniaturized primary airway epithelia and reporter viruses development to screen antiviral molecules against respiratory viruses



Authors:

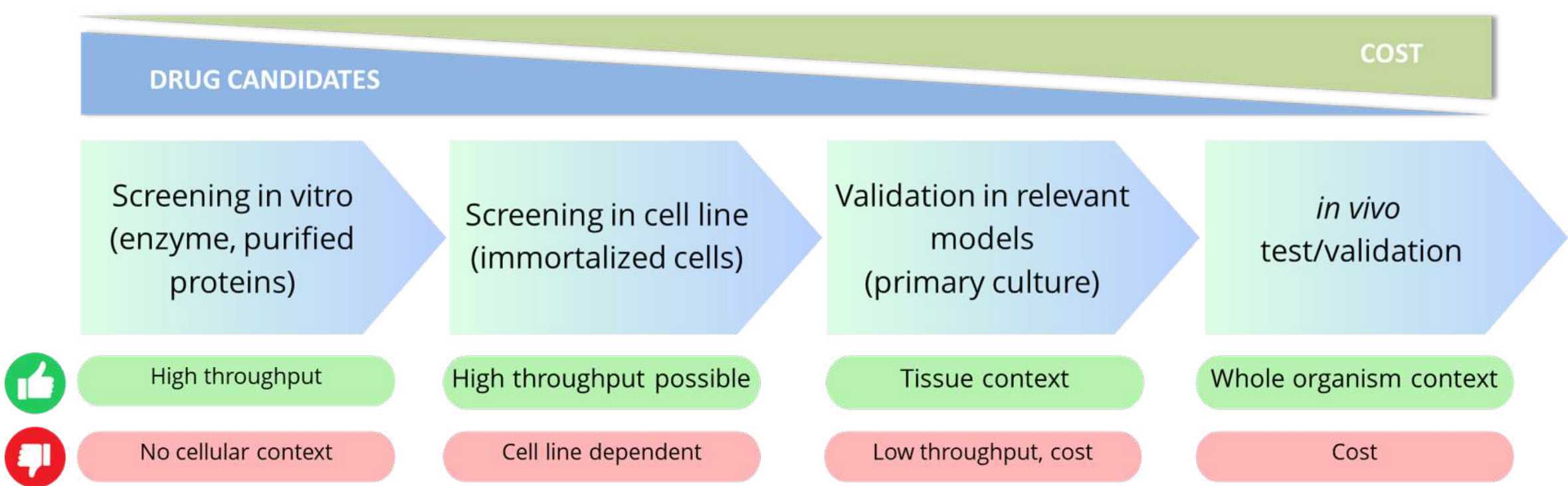
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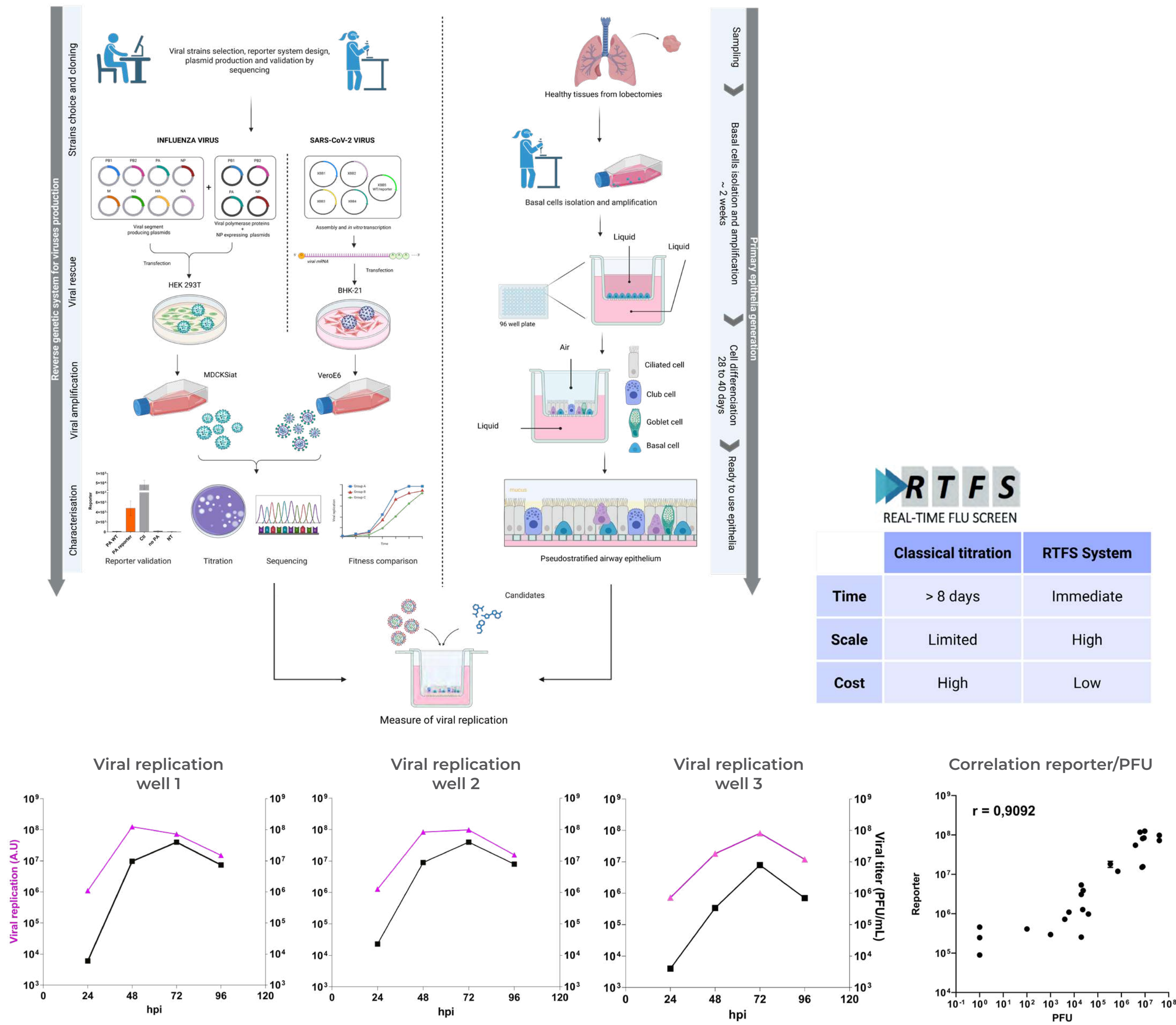
Introduction

Discovery of new antiviral drugs against respiratory viruses remains limited. The COVID-19 pandemic exposed the limitations of current screening methods, which mostly rely on immortalized cell lines and lab-adapted viral strains—failing to reflect the proper infection of human tissues. To address these issues, we are developing a more physiologically relevant screening platform using primary human airway epithelia in a 96-well air-liquid interface (ALI) format, combined with recent influenza and SARS-CoV-2 unique reporter strains. This system enables rapid, real-time, and cost-effective drug screening and evaluation. We believe that the use of a more relevant cellular model should improve the quality of results, increase the success rate *in vivo* and therefore reduce the number of animals required, in line with the 3Rs principle.

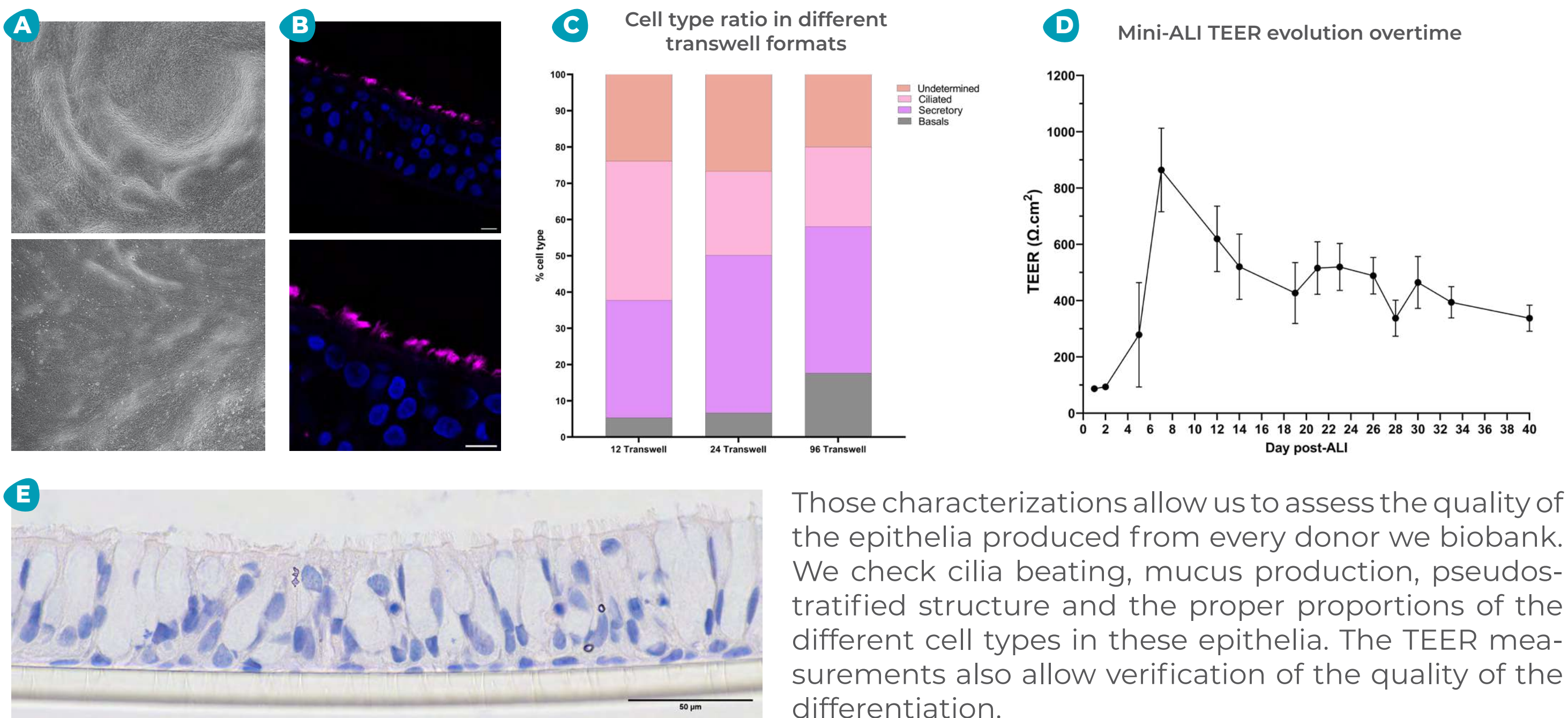
1 Drug candidate identification and validation steps



2 System set-up and readout

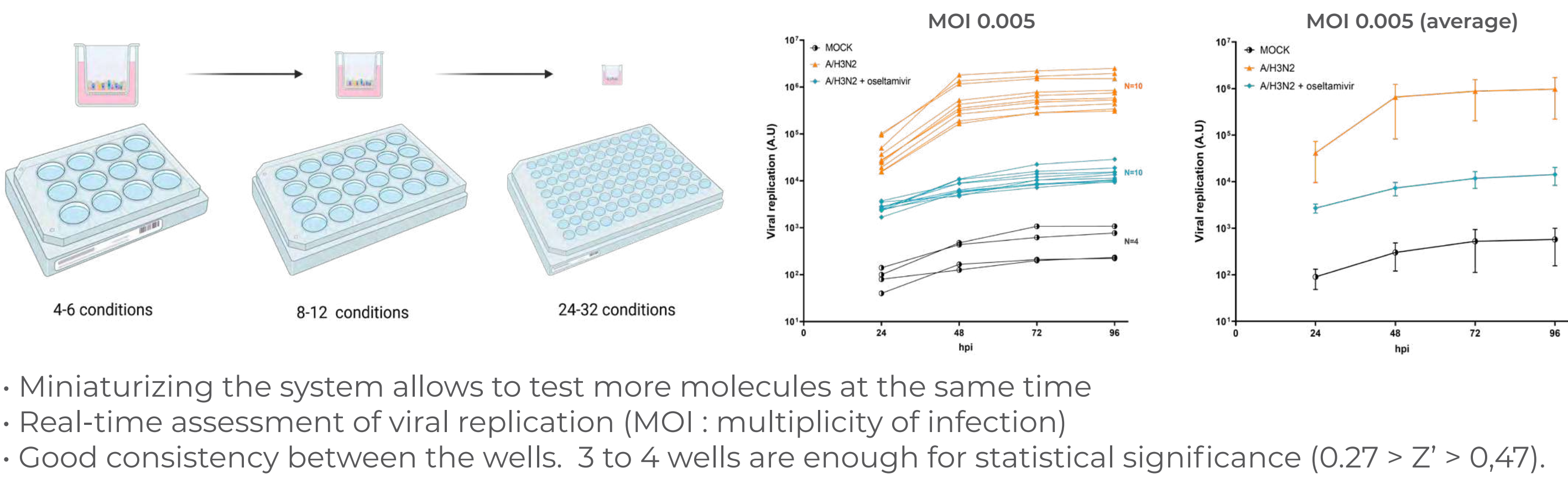


3 Human primary airway epithelium model validation

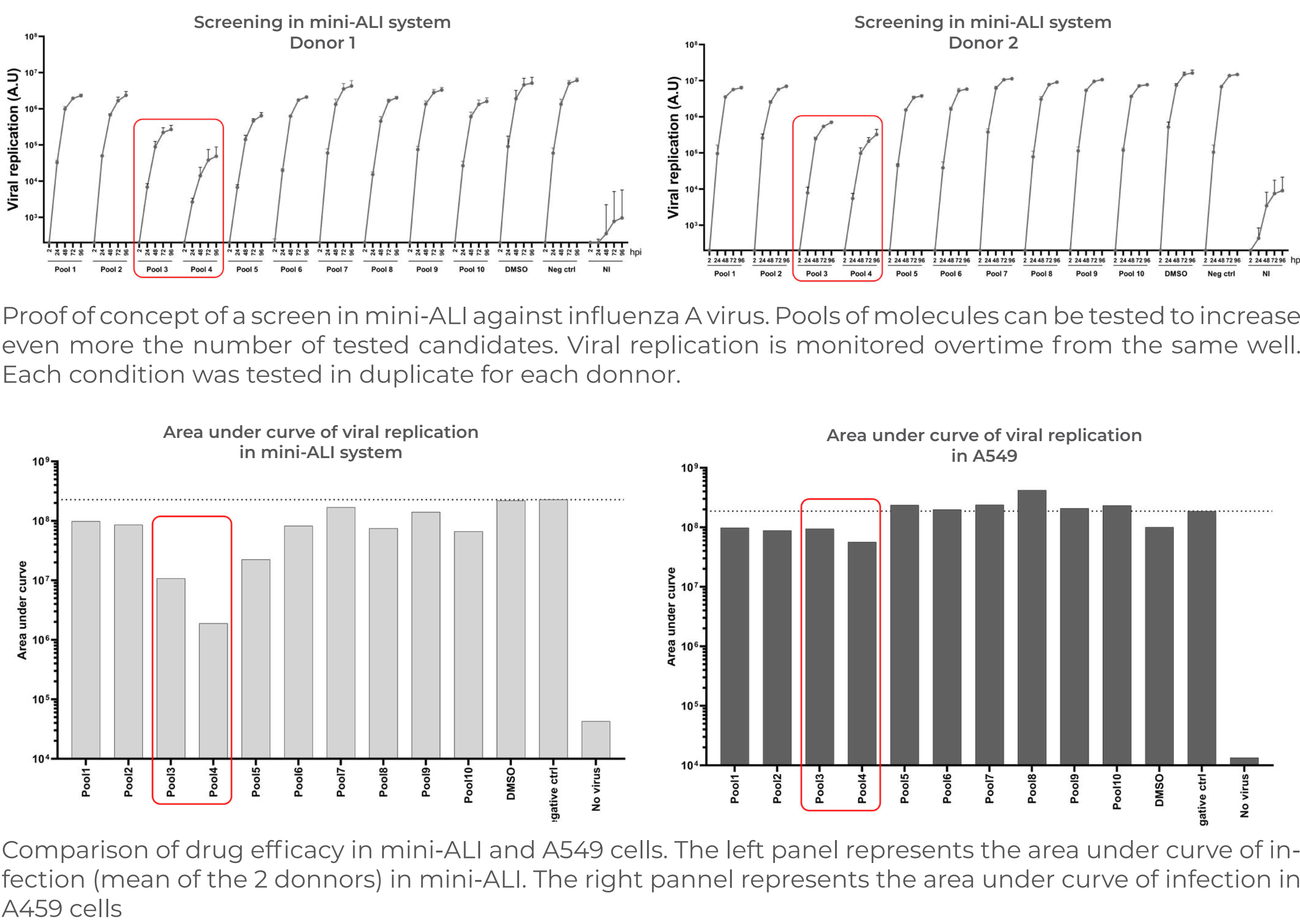


Those characterizations allow us to assess the quality of the epithelia produced from every donor we biobank. We check cilia beating, mucus production, pseudostratified structure and the proper proportions of the different cell types in these epithelia. The TEER measurements also allow verification of the quality of the differentiation.

4 System miniaturization in 96 well plate format

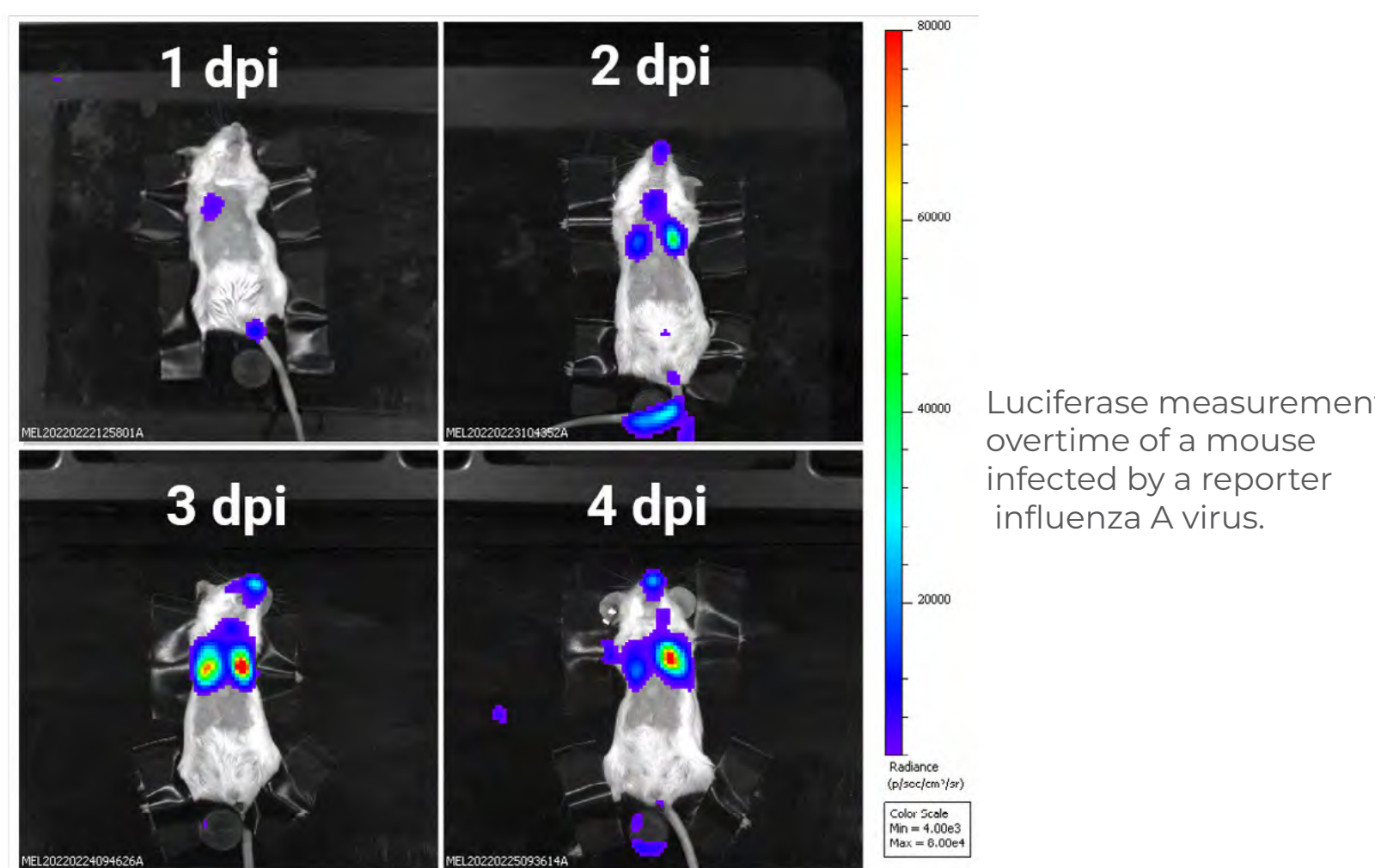


5 Compound screening in mini-ALI system and comparison with A549 cell line



6 In vivo testing of antiviral hits

Following-up the tests in primary mini-ALI system, hit candidates can be tested *in vivo* in ERBC Dommartin's facility.



Conclusion

By integrating the ALI (Air-Liquid Interface) system and recent viral strains into our advanced drug screening service, we aim to enhance the quality and relevance of antiviral hits. This approach reduces both the duration and cost of screenings. It also reduces the selection of false positives and false negatives. The use of up-to-date viral strains ensures that candidate drugs target current circulating viruses effectively. Additionally, miniaturizing the ALI system and combining it with real-time screening technology allow us to test numerous drugs simultaneously, overcoming traditional technical limitations and bottlenecks of ALI culture. As experts in *in vivo* experimentation, we can also validate the hits identified in ALI screenings directly in animal models.

